

Hydrolytic Activity of Muscle Acyl Phosphatase on 3-Phosphoglyceryl Phosphate¹

Muscle acyl phosphatase (E.C. 3.6.1.7) catalyzes the hydrolysis of many acyl phosphates^{2,3}; the action of this enzyme on physiological acyl phosphates as carbamyl phosphate⁴ and 3-phosphoglyceryl phosphate^{5,6} is of great interest. Both HARARY⁵ and KRIMSKY⁶ had, in fact, already carried out research on the hydrolysis of 3-phosphoglyceryl phosphate by acyl phosphatase prepared from muscle. HARARY's measurements were calculated indirectly, by determining the rate of P_i formation by addition of acyl phosphatase purified 600 times. KRIMSKY⁶, through a direct method, had shown the catalytic activity of acyl phosphatase on the hydrolysis of 3-phosphoglyceryl phosphate; the enzyme used, prepared according to the method of KOSHLAND⁷, had a purification factor of 37.

In this paper a direct determination of 3-phosphoglyceryl phosphate hydrolysis by a highly purified preparation of muscle acyl phosphatase is reported.

Materials and methods. Acyl phosphatase was prepared from horse muscle according to the method of GUERRITORE et al.⁸. The final preparation obtained had a specific activity of 380, expressed as μ moles of split acetyl phosphate/min/mg of protein at 25°C and pH 5.3, with a 2300-fold purification.

Glyceraldehydephosphate dehydrogenase (E.C. 1.2.1.9), alcohol dehydrogenase (E.C. 1.1.1.1), NAD and NADH₂ were purchased from C. F. Boehringer & Söhne G.m.b.H., Mannheim (Germany). 3-Phosphoglyceraldehyde was purchased from SIGMA Chem. Co. All other reagents were obtained from E. Merk AG.

3-Phosphoglyceryl phosphate was prepared according to the method of NEGELEIN and BRÖMEL⁹, slightly modified in order to avoid possible impurities of sulphate, that is an inhibitor of muscle acyl phosphatase^{6,10}. To 500 μ moles of 3-phosphoglyceraldehyde were added 1 m-mole of phosphate, 24 μ moles of NAD, 0.6 mg of glyceraldehydephosphate dehydrogenase, 3 mg of alcohol dehydrogenase and 0.5 m-moles of acetaldehyde in fractions of 0.05 m-moles. This mixture, kept to pH 7.6, was incubated for 25 min at 18°C and then acidified at pH 2.6 with perchloric acid. The product was precipitated by 500 ml of cold acetone, acidified with the same quantity of per chloric acid used to acidify the mixture. The precipitate was again washed with cold acidified acetone and dissolved in 3–4 ml of water. This solution was centrifuged to remove the enzymes. The supernatant was then brought to pH 7 by 5M K₂CO₃ and kept at –20°C. The enzymes were previously dialyzed against 0.1 M triethanolamine hydrochloride-NaOH buffer pH 7.6, containing EDTA 0.01 M.

3-Phosphoglyceryl phosphate was measured at 340 nm by NADH₂ oxidation in the presence of glyceraldehydephosphate dehydrogenase¹¹. Phosphate determination

was carried out according to FISKE and SUBBAROW¹² on 3-phosphoglyceryl phosphate. By this procedure the labile phosphate in 3-phosphoglyceryl phosphate was split off and directly determined without previous hydrolysis; in our preparation a maximal P_i impurity of about 20% could be estimated by this way. NAD and

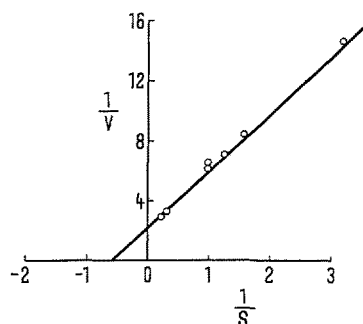


Fig. 1. The effect of 3-phosphoglyceryl phosphate concentration on acyl phosphatase activity. The data are plotted according to LINEWEAVER and BURK¹⁶. The incubation mixture at 25°C contained acetate buffer, pH 5.3, 80 μ moles; 3-phosphoglyceryl phosphate at the concentration indicated; acyl phosphatase (diluted before addition), 10 μ l; in a total volume of 1 ml. V, μ moles of substrate, hydrolyzed/min. S, substrate concentration (mM).

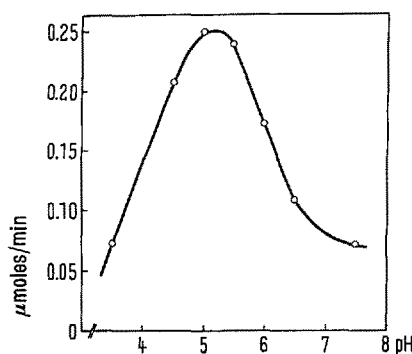


Fig. 2. Effect of pH on the 3-phosphoglyceryl phosphate splitting by acyl phosphatase. The incubation mixture at 25°C contained β - β' -dimethyl-glutaric acid-NaOH buffer, at the pH values indicated, 80 μ moles; 3-phosphoglyceryl phosphate, 2 μ moles; acylphosphatase diluted before addition, 10 μ l; in a total volume of 1 ml.

Relative rate of enzymatic hydrolysis and K_m for acetyl phosphate and 3-phosphoglyceryl phosphate as substrates of acyl phosphatase^a

| Substrate | Relative rate of enzymatic hydrolysis | K_m (mM) |
|-----------------------------|---------------------------------------|------------------|
| Acetyl phosphate | 1 | 7.7 ^b |
| 3-Phosphoglyceryl phosphate | 1.2 | 1.6 |

^a The rate with acetyl phosphate is taken as unity. ^b Datum from ⁴.

¹ This work was supported by a grant from the Impresa Enzimologia of Italian Consiglio Nazionale delle Ricerche.

² F. LIPMANN, *Adv Enzymol.* 6, 231 (1946).

³ G. RAMPONI, C. TREVES and A. GUERRITORE, *Archs Biochem. Biophys.* 115, 129 (1966).

⁴ G. RAMPONI, F. MELANI and A. GUERRITORE, *Ital J. Biochem.* 10, 189 (1961).

⁵ I. HARARY, *Biochim. biophys. Acta* 26, 434 (1957).

⁶ I. KRIMSKY, *J. biol. Chem.* 234, 228 (1959).

⁷ D. E. KOSHLAND JR., in *Methods in Enzymology* (Ed. S. COLOWICK and N. O. KAPLAN; Academic Press, New York 1955), vol. II, p. 555.

⁸ A. GUERRITORE, G. RAMPONI and V. BACCARI, 1st Meeting Abstr. of the Feder. Europ. biochem. Soc. (London 1964), p. 18.

⁹ E. NEGELEIN and H. BRÖMEL, *Biochem. Z.* 301, 135 (1939).

¹⁰ A. GUERRITORE, G. RAMPONI and A. ZANOBINI, *Boll. Soc. ital. Biol. sper.* 36, 1986 (1960).

¹¹ E. NEGELEIN and H. BRÖMEL, *Biochem. Z.* 303, 132 (1939).

¹² C. H. FISKE and Y. SUBBAROW, *J. biol. Chem.* 66, 375 (1925).

3-phosphoglycerate were also present as impurities, but their amounts, determined through optical methods¹³, were so low that they did not interfere in our experiments.

The hydrolytic activity of acyl phosphatase on 3-phosphoglyceryl phosphate was measured by the above mentioned optical test, which estimates the residual substrate after suitable periods of time. The enzymatic reaction was stopped by adding ammonium sulphate in final concentration 0.4 M: this concentration completely inhibits muscle acyl phosphatase^{6,10}. In our conditions, the estimation of enzymatic hydrolysis, by using the method of LOWRY and LOPEZ¹⁴ for the inorganic phosphate, is not suitable due to the catalytic effect of molybdate on the hydrolysis of the 1-radical of 3-phosphoglyceryl phosphate¹⁵.

Results. The Table reports the acyl phosphatase activity on both acetyl phosphate and 3-phosphoglyceryl phosphate, expressed as relative rate of enzymatic hydrolysis.

Figure 1 shows a LINEWEAVER-BURK¹⁶ plot, from which the Michaelis constant for 3-phosphoglyceryl phosphate at pH 5.3 was evaluated: this value is also reported in the Table.

It can be seen (Table) that 3-phosphoglyceryl phosphate is easily hydrolyzed by the enzyme, furthermore the hydrolysis rate is higher and the Michaelis constant is lower than with acetyl phosphate.

The effect of pH on the enzymatic hydrolysis of 3-phosphoglyceryl phosphate is reported in Figure 2. The optimum pH for acyl phosphatase activity on this substrate results in about 5.3, as previously obtained with other acyl phosphates^{3,4,17}.

These results add some quantitative information about the acyl phosphatase action on 3-phosphoglyceryl phosphate; this hydrolytic activity can explain the increase both of alcoholic fermentation⁸ and that of glycolysis^{18,19}, under particular conditions²⁰.

Riassunto. Viene studiata l'attività idrolitica dell'acilfosfatasi purificata da muscolo sull'acido 1,3-difosfoglicerico. L'ottimo di pH è uguale a quello ottenuto per altri acilfosfati. Il valore della costante di Michaelis è dello stesso ordine di grandezza di quello trovato per l'acetilfosfato.

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Istituto di Chimica biologica dell'Università di Firenze (Italy), 5 May 1967.

¹³ *Methods of Enzymatic Analysis* (Ed. H. U. BERGMAYER; Verlag Chemie Academic Press, New York and London 1963), p. 528 and p. 224.

¹⁴ O. H. LOWRY and J. A. LOPEZ, *J. biol. Chem.* **162**, 421 (1946).

¹⁵ H. WEIL-MALHERBE and R. H. GREEN, *Biochem. J.* **49**, 286 (1951).

¹⁶ H. LINEWEAVER and D. BURK, *J. Am. chem. Soc.* **56**, 658 (1934).

¹⁷ I. HARARY, *Fedn Proc. Fedn Am. Socs exp. Biol.* **16**, 192 (1957).

¹⁸ V. BACCARI, A. GUERRITORE, G. RAMPONI and M. P. SABATELLI, *Boll. Soc. ital Biol. sper.* **36**, 360 (1960).

¹⁹ I. HARARY, *Fedn Proc. Fedn Am. Socs exp. Biol.* **21**, 87 (1962).

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Abnormalities of the Eye Pigments (Pteridins and Ommochromes) Induced in *Drosophila melanogaster* by the Inhibitor of Xanthine Dehydrogenase 4-Hydroxypyrazolo (3,4 d) Pyrimidine

When wild-type strains of *Drosophila melanogaster* are grown on media containing the inhibitor 4-hydroxypyrazolo (3,4 d) pyrimidine (HPP), phenocopies are obtained which mimic the *ma-l* and *ry* mutants. In fact, loss of isoxanthopterin and uric acid with accumulation of the corresponding precursors 2-amino-4-hydroxypteridine and hypoxanthine, as well as diminution of the red pteridins of the eye, have been observed^{1,2}. These phenomena do not exclude the possibility that the inhibitor might affect other mutants, with different patterns of eye pigments metabolism.

Strains of the *cl* and *se* mutants were raised, according to KELLER and GLASSMAN¹, on different concentrations of HPP: 0.0; 0.01; 0.02; 0.03; 0.04; 0.06; 0.08 and 0.1 g%. Eye colours were recorded daily when the adults emerged. Flies, 3–5 days old, were placed in boiling water for 1 min, the heads dissected and homogenized in AEA (30% ethanol and HCl q.s. to pH 2⁴), 0.01 ml for each head. Aliquots of 0.01 ml of the homogenates were laid down on Whatman No. 1 filter paper; the chromatograms, developed by ascending chromatography in *n*-propanol and 2% ammonium acetate in water (1:1) and also in 3% NH₄Cl in water⁵, were dried and observed in the visible light and under an UV-lamp emitting mainly at 365 nm.

The remaining portions of the homogenates were centrifuged (20,000 g/30 min); the limpid supernatants, quantitatively collected, were diluted 1:10 and the absorption was measured with a Beckman DU spectrophotometer. The bodies also were treated in the same manner.

In the *cl* mutant, grown on HPP, the eye colour remains dark maroon. Loss of isoxanthopterin and accumulation of 2-amino-4-hydroxypteridin, mainly in the bodies of the male flies, occur at about the same concentrations of HPP as in wild-type strains. Partial loss of the drosoppterins is more prominent for neodrosoppterin than for iso- and drosoppterin; correspondingly accumulation of bioppterin, but not of sepiapterins, could be observed.

In *se* strains, grown on HPP (more evidently at the higher concentrations) the eyes are more pale, having a brown-beige colour, and usually smaller as compared with control flies. Loss of isoxanthopterin is in *se*, as in wild-strains, almost complete at the concentrations of HPP higher than 0.04%; accumulation of 2-amino-4-hydroxypteridine seems to be present in somewhat

¹ E. C. KELLER and E. GLASSMAN, *Nature* **208**, 202 (1965).

² P. BONI, B. DE LERMA and G. PARISI, *Experientia* **23**, 186 (1967).

³ We are indebted to 'Wellcome Italia S.p.A.', Rome, for a generous gift of HPP (Allopurinol®).

⁴ B. EPHRUSSI and J. L. HEROLD, *Genetics*, Princeton **29**, 148 (1944).

⁵ M. VISCONTINI, E. HADORN and P. KARRER, *Helv. chim. Acta* **40**, 579 (1957).